

---

## Correlation between Density of *Vibrio* Bacteria with *Oscillatoria* sp. Abundance on Intensive *Litopenaeus vannamei* Shrimp Ponds

Heri Ariadi, Mohammad Mahmudi, Mohamad Fadjar

Fisheries and Marine Science Faculty, University of Brawijaya, Indonesia

Email address: [ariadi\\_heri@yahoo.com](mailto:ariadi_heri@yahoo.com)

---

### KEYWORDS

Intensive ponds  
*Litopenaeus*  
*vannamei*  
*Oscillatoria* sp  
*Vibrio* bacteria

**Abstract** The abundance of *Vibrio* bacteria and the presence of an excess *Oscillatoria* sp plankton type are the main problems that often arise in intensive vanamei (*Litopenaeus vannamei*) shrimp culture. To examine this problem, the purpose of this study was to determine of model approach interaction between the abundance of *Vibrio* and *Oscillatoria* sp with the shrimp growth rate in ponds. This research was conducted with the ex-*post facto* design on intensive shrimp culture operations in Bayeman Village, Probolinggo. The results data from field research variable are analyzed using a dynamic modeling system. From the modeling analysis results, showed that the *Oscillatoria* sp abundance pattern tended to increase over with shrimp culture period as following the pattern of tropical status dynamics by the nutrient increase load of 16.37%/week in ponds ecosystem. Meanwhile, the increase of *Vibrio* colonies density in ponds continued to increase aggregately by 0.99%/week on nine weeks and 12.5%/week on the last eight weeks of shrimp culture periods. So, it can be concluded that the fluctuations density of *vibrio* and *Oscillatoria* sp bacteria in ponds is are bioecological responses from increased nutrient loads and other micromaterials in ponds due to the longer period of shrimp culture.

---

### Introduction

*Litopenaeus vannamei* shrimp culture is the strategic pillars from fisheries production commodities that are very promising in several tropical Asian countries (Ashton, 2007 and Syah, 2017). Since it was introduced and developed in Indonesia from at 2000s decade, the production and development of vanname shrimp on aquaculture activities have continued to increased (Romadhona *et al*, 2016 and Yi *et al*, 2016). Data from the report by Directorate General of Aquaculture (2016), the last production of vanname shrimp in Indonesian reached 488,019 tons and is predicted to increase at 10% per year.

On the shrimp culture bussines, increase of shrimp harvest productivity due to intensive on shrimp culture system, has a negative impact on the increasing prevalence of spreading germs,

which is caused by increasing cultivation waste (Jayanthi *et al*, 2018 and Mohan *et al*, 2019). This is usually characterized by a decrease in the quality standards of pond water quality. Water quality parameters are vital components that should role playing in the viability of vannamei shrimp culture (Rahman *et al.*, 2015 and Zafar *et al.*, 2015). some variables of water quality that have a significant influence to ecosystem dynamics in vannamei shrimp farming ponds are the presence of *Blue Green Algae* (BGA) plankton and uncontrolled abundance of *vibrio* bacteria (Boyd and Tucker, 1998; Weidemann, 2002; Cardenas *et al.*, 2017; and Stalin *et al.*, 2017).

*Blue Green Algae* (BGA) plankton or cyanobacteria such as *Oscillatoria* sp in the event of blooming will cause shrimp to become off-flavour due to high levels of geosmine

(Lovell *et al.*, 1985 and Tucker, 2000). Whereas vibrio is a type of pathogenic bacteria that is the cause of various serious problems in aquaculture (Chatterjee and Haldar, 2012). From the description of the problem, the purpose of this study was to determine interaction model between the abundance of vibrio bacteria with presence of *Oscillatoria sp* plankton on the growth rate of *Litopenaeus vannamei* shrimp in intensive pond culture.

### Materials and methods

This research was conducted on intensive ponds in Bayaman Village, Tongas District, Probolinggo Regency used by *ex post facto* design research on a 400 m<sup>2</sup> vannamei shrimp pond during the operational cycle period of shrimp culture or precisely from July to November 2018. structured once every week from the start post until total harvest periods. So, the data will be collected according to the time series of the cultivation age. The research variables measured were water quality parameters which included TAN, NO<sub>2</sub>, TOM, temperature, plankton density, and total vibrio bacteria. While the shrimp biological parameters observed were average body weight (ABW) and average daily gain (ADG) which were determined based on periodic sampling.

The method measurement of the concentration TAN and NO<sub>2</sub> parameters was determined by spectrophotometry methods, while measurement of TOM parameters was determined by titrimetry methods at analyzed in Training Center Laboratory CP Prima Company, Paiton, Probolinggo. And all of them refer to applying procedure by APHA (1980), while temperature parameters are measured *in situ* on ponds using a Hg Thermometer.

*Vibrio* bacterial samples were analyzed in the Training Center Laboratory. CP Prima Company, Paiton, Probolinggo. Bacterial samples from the pond were taken as much as

50 ml, then before planting in media, a gradual dilution was carried out (up to 10<sup>-2</sup>), and planting was done on TCBS media so that in the petri disk with spread method. After that incubation at room temperature for 24 hours. Then the next step is to counted at the number of TVC (Total Vibrio Count) colonies prefer by method of Prescott *et al.* (2002).

Analyzed the density and diversity of plankton species were calculated and identified using hemocytometer microscopically with Olympus Type cx22 microscope based applied procedure by APHA (1980). Then plankton abundance is calculated by the formula:

$$\sum \text{cell/ml} = N \times 10^{-1} / 1 \times 10^4 \text{ cm}^3$$

Where: N is the number of plankton calculated, 10<sup>-1</sup> is the value of the diluent factor, and 1 x 10<sup>4</sup> cm<sup>3</sup> is the size of the volume in the hemocytometer box.

To determine the diversity of composition and number individual plankton in the pond, it was determined using the Shannon-Wiener diversity index (Michael, 1995) using the formula:

$$H' = -\sum p_i \ln p_i; p_i = n_i/N$$

Where: H' is the value of the diversity index, p<sub>i</sub> is the number of individuals of type i (n<sub>i</sub>) divided by the total number of individuals in the sample (N).

The technique used for collect growth parameters of shrimp is done by random sampling using nets then weighed from several pond points to obtain shrimp body weight (ABW), then to determine the level of growth rate (ADG) of shrimp then calculated using formula by Jiao *et al.* (2014) as follows:

$$\text{ADG} = \frac{\text{ABW actual sampling} - \text{ABW after sampling}}{\text{sampling periode}}$$

Then, to construct an interaction model between the abundance of vibrio bacteria with presence of *Oscillatoria sp* on the shrimp growth rate, it was processed into a modeling analysis with Stella<sup>TM</sup> software Ver. 9.12.

## Results and discussions

### Profile of Ponds Culture

The operational cycle of shrimp culture in the implementation of this study lasted 85 days. From the process of shrimp culture periods, the profile of the average shrimp growth rate (ADG) was obtained as an indicator of growth and

water quality variables which included the average concentration of nitrite parameters ( $\text{NO}_2$ ), Total Ammonia Nitrogen (TAN), temperature, total vibrio, total organic matter (TOM), as well as the Blue Green Algae abundance of type *Oscillatoria sp.* as an indicator of pond environment (Table 1).

**Table 1. Average Profile of Water Quality and Shrimp Growth During One Cycle.**

DOC (day)	ADG (gr)	Feed/day (kg)	$\text{NO}_2$ (mg/L)	TAN (mg/L)	Temperature (°C)	TOM (mg/L)	TVC (CFU/ml)	<i>Oscillatoria sp zbun</i> (cell/ml)
85	0.27	53.58	0.02	0.140	29.3	113.51	5.87E+02	4.69E+05

The value of average daily gain (ADG) by shrimp on the study shows that shrimp growth rates are running normally with weight gain per day at 0.27 gr, this result is better than results by Junda (2018) which gets an ADG value 0, 26 in intensive vanname shrimp ponds during 112 days. The value of the water quality variable for the nitrite ( $\text{NO}_2$ ) parameter of 0.02 mg/L detected is still low compared to the results by Rochin *et al.* (2017) study of 0.28-0.62 mg/L which is also a toxic level by the compound nitrite. Likewise, the TAN value, which is calculated to be 0.140 mg/L, is still lower than the results of previous studies by Gross *et al.* (2004) of 0.2 mg/L. Temperature parameters measured at 29.3°C are the optimal range of parameters for the growth of vanname shrimp cultivation (Singh *et al.*, 2013). While high levels of organic matter are caused by input feed input and fertilization that continues to grow throughout the cultivation period (Schober *et al.*, 2007).

The abundance of vibrio bacteria in ponds research average still tends to be lower ( $10^2$  CFU/ml) than compared with the abundance vibrio bacteria results of detected on intensive shrimp ponds in India by Gopal *et al.* (2005), which obtained an abundance of vibrio bacteria on an average at  $10^4$  CFU/ml from 15 pond sample points on the west and east coasts of India. The level of vibrio pathogenicity and abundance in ponds is strongly influenced by conditions of pond water quality parameters, such as temperature and dissolved oxygen concentration (Orozco *et al.*, 2007). While the high density of *Oscillatoria sp* plankton which is a Blue Green Algae species caused by the condition from pond water parameters that tend to be eutrophic or even hyper-eutrophic. In addition, the type of *Oscillatoria sp* can be fix capture Nitrogen from diffusion so that the population will be easy to Bloom (Aliviyanti *et al.*, 2017).

### Water Quality Characteristic

#### The Concentration of TAN and Nitrite

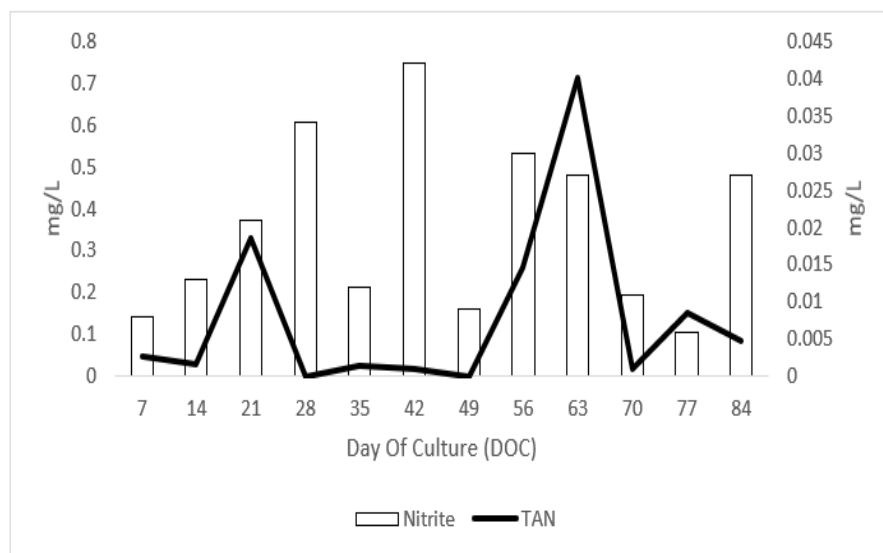
TAN concentration during culture periods ranged from 0.000-0.713 mg/L while Nitrite ( $\text{NO}_2$ ) concentrations ranged from 0.006-0.042 mg/L (Figure 1). The standard optimum of TAN and Nitrite ( $\text{NO}_2$ ) concentrations according to Edhy *et al.* (2010) is 0.10 mg/L and 0.2 mg/L. The highest increase TAN concentration occurred at 63 days with 0.713 mg/L and the lowest at 28 and 49 days with 0.0 mg/L. Because at 28 days periods can be siphoning activity was first performed after the period of *blind feeding* shrimp farming and at the age 49 days there was a recirculation activity. While the maximum concentration of Nitrite ( $\text{NO}_2$ ) occurred at 42 days by 0.042 mg/L and the lowest at 77 days by 0.006 mg/L. This is suspected on 42 days culture periods is a biological synthesis of changes

in Ammonia ( $\text{NH}_3$ ) compounds into Nitrite ( $\text{NO}_2$ ) by *Nitrosomonas sp* strain bacteria intens on the nitrification cycle.

While the decline minimum value at 77 days culture is caused by a decrease amount of feed given at that periods. High TAN concentrations are caused by increased management and increasing frequency of feeding with poor management of pond bottom sludge (Hopkins *et al.*, 1994 and Jescovitch *et al.*, 2017). Variations of TAN and  $\text{NO}_2$  concentration can also be due to the fertilization process. Fertilizing activities can increase the level of inorganic nutrient on pond ecosystems (Suwanpakdee *et al.*, 2010).

The level of concentration TAN and Nitrite ( $\text{NO}_2$ ) in the pond ecosystem, naturally is also

influenced by the effectiveness of the nitrification cycle that is taking place. Nitrification is part of the biogeochemical cycle of nitrogen changes in aquaculture ecosystems that remodel ammonia ( $\text{NH}_3$ -) to Nitrite ( $\text{NO}_2$ ) by *Nitrosomonas sp* bacteria and convert of Nitrite ( $\text{NO}_2$ ) to Nitrate ( $\text{NO}_3$ ) by *Nitrobacter* (Hastuti, 2011). Nitrogen biogeochemical processes in shrimp farming which are dominated from nitrification cycle, besides being determined by nitrifying bacteria activities are also influenced of water quality stability conditions and the use of paddle wheels aerators as aquaculture engineers (Hargreaves, 1998 and Fernandes *et al.*, 2010).



**Figure 1.** Concentration of TAN (Total Ammonia Nitrogen) and Nitrite ( $\text{NO}_2$ ) in Ponds.

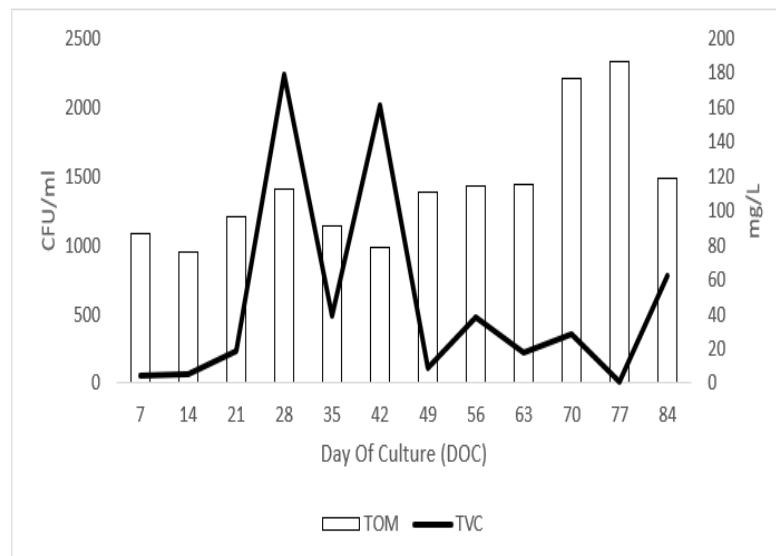
The proportion of TAN and Nitrite ( $\text{NO}_2$ ) levels will increase with pH and temperature conditions. High TAN and Nitrite ( $\text{NO}_2$ ) concentrations in water will affect the permeability of body cells on aquatic organisms and reduce of the ions concentration level (Edhy *et al.*, 2010). To manage so that the TAN and Nitrite ( $\text{NO}_2$ ) parameters in the farm remain below the threshold and are not toxic, it can be used to add probiotic treatment and maintain the stability of daily fluctuations in water quality parameters such as pH, dissolved oxygen, temperature, and salinity (Nurhatijah *et al.*, 2016 and Jaganmohan *et al.*, 2018).

#### *Organic Materials and Vibrio Bacteria*

Organic matter concentration in the ponds during intensive shrimp culture periods measured by parameters of Total Organic Matter (TOM) ranged from 76.07-186.50 mg/L. While density vibrio

bacteria were measured by the abundance of Total Vibrio Count (TVC) on TCBS media. The count was calculated to be between 50-2,230 CFU/ml (Figure 2). The optimum standards for organic matter on intensive ponds and the abundance of vibrio bacteria according to regulation by the Minister of Marine and Fisheries of the Republic Indonesia No. 75 tahun 2016, concerning General Guidelines for Enlargement of *Penaeus monodon* and *Litopenaeus vannamei* shrimp is  $\leq 90$  mg/L for TOM concentration and  $\leq 1.000$  CFU/ml for abundance vibrio bacteria. The highest concentration of organic matter occurs on 77 days shrimp culture at 186.50 mg/L, this is thought to be caused by the amount of organic material such as particle suspension, turbidity level, and accumulation of material that settles in the bottom sediment pond which is increasing due to the old cultivation periods.

While the lowest levels of organic matter were detected on 14 days culture periods at 76.07 mg/L, the phenomenon seemed reasonable because at these conditions it was the initial period of maintenance so that addition of cultivation inputs was still considered small. Meanwhile, the highest vibrio bacterial density was observed at 28 days as many as 2.230 CFU/ml.



**Figure 2.** Concentration of TOM (Total Organic Matter) and Vibrio Abundance in Ponds

This result was suspected because at that age it was a *blind feeding* period so that there was no control over feed inputs, so that it allegedly had an indirect impact on increasing levels of waste load in the ecosystem pond. Meanwhile, the lowest density of vibrio bacteria at the 77 days culture periods is 10 CFU/ml, which is possible because at that periods there has been a decrease in the number of inputs to aquaculture ponds. The high concentration of organic matter and the density of vibrio bacteria that are

pathogenic in pond ecosystems is an indication of the accumulation of waste loads due to aquaculture activities, which are usually caused by increased feed inputs, phytoplankton blooms, poor water quality management practice and unsupportive to environmental factors (Lekshmi *et al.*, 2014). Vibrio bacteria that are sensitive to environmental conditions will be very opportunistic if their abundance is above the threshold (Heenatigala and Fernando, 2016). So that, it will cause various multiple diseases in cultivated shrimp such as

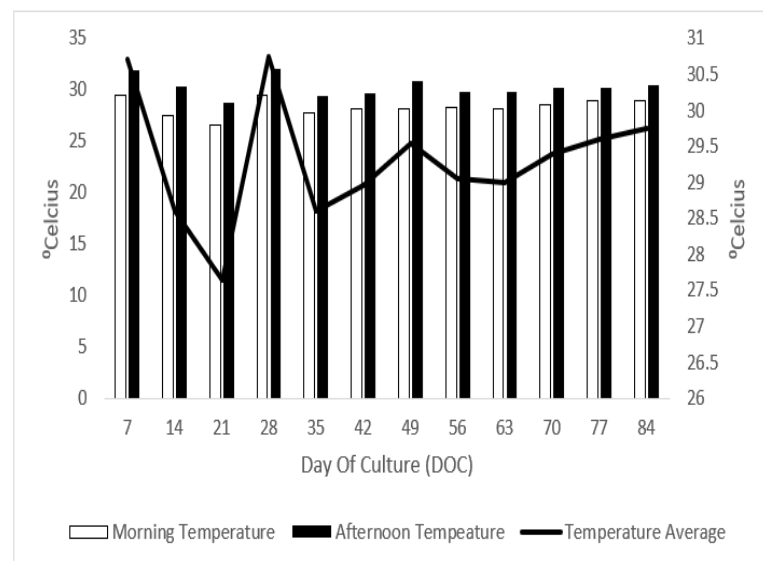
white feces syndrome, *Enterocytozoon hepatopenaei* (EHP) infection, and mass death caused by physiological symptoms caused by *Vibrio* bacterial infections (Jayasree *et al.*, 2006; Sriurairatana *et al.*, 2014; Mastan, 2015; and Aranguren *et al.*, 2017).

From Figure 2, it can be seen that the distribution of vibrio bacteria decreases after 42 days shrimp periods, while the level of organic matter in ponds ecosystem shows a graph of the increase in 42 days shrimp periods. It can be synthesized that at the 42 days culture periods there was a dominance competition between vibrio and plankton due to bioaugmentation through periodic treatment by probiotics bacteria. The bioaugmentation process will be beneficial in maintaining the stability of water quality and suppressing the presence of pathogenic bacteria without having to reduce the ratio nutrients for phytoplankton (Janeo *et al.*, 2009). In addition, the decrease of vibrio

bacteria density of at the age of 42 days it can also due to other factors such as the presence of water circulation, effective feed management, and stabilizing pond alkalinity levels (Mangampa, 2015).

#### Daily of Temperature Fluctuations

Temperature is a water quality parameter that plays an important role in the course of the vannamee shrimp culture system (Abdelrahman *et al.*, 2018). In ponds culture, the level of temperature is strongly influenced by weather and environmental conditions (Culberson and Piedrahita, 1996). In this study, the temperature level in the morning ranged from 26.6-29°C with an average of 28.37°C, while during the day it ranged from 28.7-32°C with an average temperature at 30.28°C and overall average temperature values in the farm range from 29.3°C (Figure 3).



**Figure 3.** Level of Fluctuations and Average Temperature Daily in Ponds.

In the vannamee shrimp culture, the level of temperature fluctuations greatly affects the metabolic rate and the physiological response of shrimp to the environment as well as the decomposition process of organic matter by aquatic microorganisms (Chakravarty *et al.*, 2016). In addition, temperature fluctuation effects also affect to dynamics of water quality parameters and the dominance composition of plankton in ponds (Palafox *et al.*, 2019). The temperature values in farms vary depending on the shape of the region's topography and weather conditions (Islam *et al.*, 2004).

The close relationship between temperature and weather conditions will indirectly have an impact on the productivity of shrimp harvest (Rimi *et al.*, 2013). Due to unpredictable weather changes will increase vulnerability and poor adaptation of shrimp to erratic environmental conditions (Soto *et al.*, 2018). To keep the shrimp from being stressed due to temperature fluctuation effect, farmers will usually grow phytoplankton to a density of  $10^6$  cell/ml in hopes of absorbing sunlight radiation during the day, so that at night the pond water conditions remain warm (Edhy *et al.*, 2010).

#### Dynamics of *Oscillatoria* sp Abundance

Plankton *Oscillatoria* sp is a type of cyanobacteria that cause HABs in aquatic ecosystems (Deep *et al.*, 2013). *Oscillatoria* sp on aquatic environment will produce toxins such as neurotoxin, anatoxin, and hepatotoxins which cause shrimp to become *off-flavour* (Smith, 1996; Rodgers, 2008; and Tho *et al.*, 2012). *Oscillatoria* sp and several other types of Blue Green Algae plankton can grow with nutrient-poor in pond waters because *Oscillatoria* sp has buoyancy and heterocyst cells that can take free nutrients from the air (Edhy *et al.*, 2010). From the research data, it shows that the abundance of *Oscillatoria* sp varies during the period of shrimp culture, with an abundance of at least 0 cell/ml at the beginning of the cultivation period and the most  $1.38 \times 10^6$  seen at 21 days cultured. It is possible that age there is no activity siphon and cultivation is still dependent on *blind feeding* system, so it is very possible to occurs nutrient imbalance in shrimp pond ecosystem. *Oscillatoria* sp abundance always exists from DOC 14 days until the harvest. While plankton abundance in ponds also varies from DOC 7 days to harvesting with the lowest amount of  $2.5 \times 10^3$  cell/ml (Figure 4).

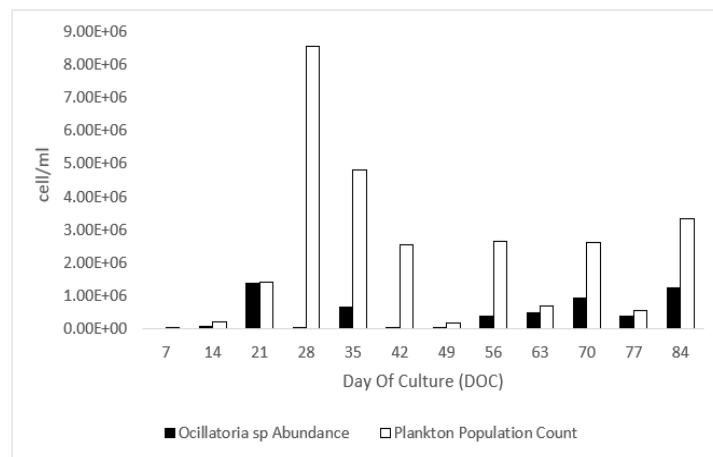


Figure 4. Plankton Populations and Existence Dynamics of *Oscillatoria* sp in Ponds.

*Oscillatoria* sp and cyanophyta are harmful algae species and mostly avoided in intensive vanname shrimp culture (Yusoff *et al.*, 2001 and Jia *et al.*, 2014). *Oscillatoria* sp and cyanophyta species that cause mud smell shrimp can grow at a ratio of N: P 1:15 or below 1:20 (Aziz *et al.*, 2015). So, it is desirable to manage N and P levels in pond waters as a limiting factor for the growth

of *Oscillatoria* sp (Xu *et al.*, 2010). In addition, the abundance and dominance of certain plankton type in the waters are also influenced by environmental physical-chemical factors such as temperature and dissolved oxygen (Bassat, 2008 and Palafox *et al.*, 2010). To overcome the *blooming* of *Oscillatoria* sp or other types of Blue Green Algae, can be used chemical materials

such as  $\text{CuSO}_4$ , Simazine, and potassium ricinoleate (Supono, 2015). Option other without chemical materials, by changing water, minimizing of nutrient input to the pond, and mixing through with aeration mechanism (Rodgers, 2008).

#### *Model Interaction Between Oscillatoria sp with Vibrio Bacteria Abundance and Shrimp Growth*

The model interaction between the existence of *Oscillatoria sp* plankton and abundance of *Vibrio sp* bacteria and its correlation to the shrimp growth rate of ponds can be seen in Figure 5.A. From interaction model that occurs of pond ecosystem, it can be interpreted based on the model validation results in Figure 5.B that an increase of shrimp biomass in ponds estimated at 6.45% per week will affect of aquatic tropics status levels to increased nutrient load. Where in this study estimated at 16.37% from an increase nutrient load on the pond per week. The increase nutrient load in intensive ponds is naturally caused by an increasing amount from a feed that continues to grow in line with the increase of shrimp growth rate (Thakur and Lin, 2003).

Changes of trophic status levels in pond aquaculture ecosystems will lead to dynamic conditions in some parameters of the shrimp culture indicator. In this study, it was shown that changes in the status tropic waters correlated positively with the increasing abundance of *Oscillatoria sp*. as seen in Figure 5.B *Oscillatoria sp*. is a cyanobacteria strain that is very tolerant from fluctuations of changes on aquatic tropics levels (Aliviyanti et al., 2017).

During the operational cycle of shrimp culture, the abundance of *vibrio* bacteria continues to increase at aggregative and temporal. Starting from the initial 3.75 weeks of shrimp culture periods there is a mild increase in the number of *Vibrio* populations or as much as 6.20% per week. And it increased to 12.12% per week in the last 8.25 weeks of shrimp culture periods. eventuality, an increase in *vibrio*

abundance is caused by the longer age of shrimp culture. So that, it will produce a lot of organic material load waste on the pond ecosystems (Nimrat et al, 2008). As well as the dynamic fluctuations in water quality throughout the shrimp culture cycle, it also contributes to the level of pathogenicity and development of *Vibrio* bacteria in ponds (Kopprio et al., 2017).

The *Oscillatoria sp* abundance can be said, that the index of growth in the pond is very closely influenced by the level of fertility on the pond ecosystem. This is in according with Pirzan and Masak (2007) who said that in natural conditions, an increase in primary productivity in pond aquaculture would be followed by an increase in the diversity of plankton such as cyanophyta and chlorophyta. The graph of the *Vibrio sp* bacteria increased from the results of conceptual validation models has an aggregative pattern of increased rate following the length of the shrimp culture periods. In intensive shrimp ponds with high stocking densities, *Vibrio sp* is often found with an average abundance between  $10^1$ - $10^3$  CFU/ml and depends on the condition of the pond waters (Tompo, 2016).

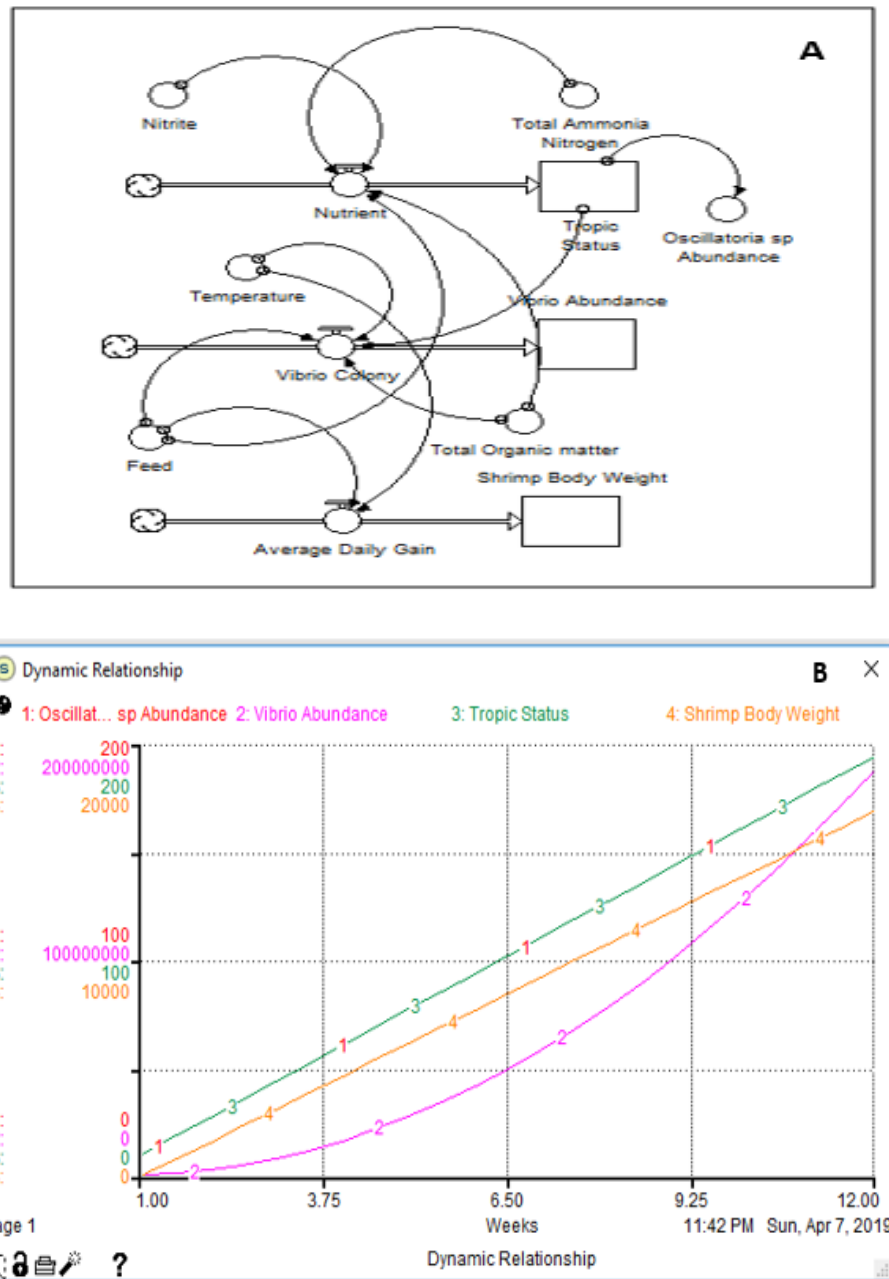
So that, it can be analyzed from the *Vibrio sp* and *Oscillatoria sp* abundance on intensive ponds is very much influenced by the increasing of aquaculture inputs and daily fluctuations of water quality (Figure 5.B), where the number and concentration continue to increase and dynamically respond to the aquaculture operational period. from increasing shrimp biomass and growing shrimp culture periods. In addition, the presence of *Oscillatoria sp* can be used as a biological parameter for the determined of status tropics in aquatic ecosystem, because accorded of biological analyzed, cyanobacteria strains can be used as implications for bioreporters or *in situ* bioindicators in aquatic environment (Mateo et al., 2015).

*Oscillatoria sp* is a Cyanophyta strain that is cosmopolite or easily develops and adapts



moderately to intensive ponds with pH, salinity, dissolved oxygen and varying temperatures (Utojo, 2015 and Harris, 2016). The *Oscillatoria* sp abundance and other types of Cyanophyta plankton will continue to tend to increase with addition age of shrimp culture periods and correlate very closely with phosphate concentration, turbidity, and salinity in ponds

(Widigdo and Wardiatno, 2013). Increased diversity of Cyanophyta and Chlorophyta types of plankton which usually tends to be followed by an increase of primary productivity in pond waters. Making these conditions, sometimes will have a negative impact both ecology, economy or environmental health (Tarunamulia *et al.*, 2016).



**Figure 5.** Model of the Relationship Between *Vibrio* sp, *Oscillatoria* sp, and Shrimp Growth by Stella™ Ver. 9.12 (A) Model Conceptual, (B) Model Validation

Whereas *Vibrio* sp is a type of gram-negative bacteria that is opportunistic pathogenic towards vannamei shrimp (Cardenas *et al.*, 2017). Shrimp infected with clinical vibriosis is characterized by the red carapace, melanosis of the skin, necrosis on tail and hepatopancreas are dark red and lesions are found (Sarjito *et al.*, 2015 and Han *et al.*, 2018). The physical-chemical quality factor of poor water is the main trigger for the vibrio abundance in intensive ponds (Kharisma and Manan, 2012). Management of handling poor water quality is the main reason why the existence of vibrio is so diverse and abundant in intensive aquaculture waters (Bintari *et al.*, 2016 and Utami *et al.*, 2016). To overcome the level of pathogenicity vibriosis on ponds, it can be carried out both chemically, physically, and biologically according to environmental conditions, culture methods and financial capabilities (Hatmanti, 2003).

Specific actually, an increasing amount of *Vibrio* bacteria abundance at week 3.75 and *Oscillatoria* sp aggregate (Figure 5.B) can be overcome by pond habitat manipulating with maintaining the stability of C : N : P Ratio. Increased and stabilizing of levels by C : N : P Ratio will increase the growth of heterotrophic bacteria and regulate the dominance of plankton communities and then to reduce of the inorganic nitrogen concentration in ponds (Masithah *et al.*, 2016). C:N ratio strategy is the most potential method for growing heterotrophic bacterial communities that are beneficial in aquaculture systems (Avnimelech, 1999). The optimum ratio of C : N to maintain for the stability of water quality and shrimp farming productivity is 15 (C) : 1 (N) (Panjaitan, 2011). While adjusting of N: P ratio according to the pond ecosystem will determine for type and plankton abundance that exist in ponds (Daruti *et al.*, 2017). The ratio optimum for N : P in shrimp culture is 16 (N) : 1(P) (Redfield *et al.*, 1963).

## Conclusions and suggestions

The conclusions from the results of this study indicate that the water quality dynamic experiences fluctuating levels of concentration over 85 days shrimp culture periods of shrimp maintenance. Whereas, the relationship between the existence and abundance of *Oscillatoria* sp and vibrio bacteria in ponds is strongly influenced by the longer age shrimp culture periods.

## References

- Abdelrahman, H. A., Abebe, A., and Boyd, C. E. 2018. Influence of Variation in Water Temperature on Survival, Growth and Yield of Pacific White Shrimp *Litopenaeus vannamei* in Inland Ponds for Low-Salinity Culture. *Aquaculture Research*, 50: 658-672.
- Aliviayanti, D., Suharjono., and Retnaningdyah, C. 2017. Cyanobacteria Community Dynamics and Trophic Status of Intensive Shrimp (*Litopenaeus vannamei*) Farming Pond in Situbondo, East Java Indonesia. *Journal of Tropical Life Science*, 7(3): 251-257.
- American Public Health Association (APHA). 1980. American Public Health Association Standart Methods for The Examination of Water and Wastewater, 15<sup>th</sup> ed. APHA/WWA-WPCT, Washington DC, USA: 1134.
- Aranguren, L. F., Han, J. E., and Tang K. F. J. 2017. *Enterocytozoon hepatopenaei* (EHP) is A Risk Factor for Acute Hepatopancreatic Necrosis Disease (AHPND) and Septic Hepatopancreatic Necrosis (SHPN) in The Pacific White Shrimp *Penaeus vannamei*. *Aquaculture*, 471: 37-42.

- Ashton, E. C. 2007. The Impact of Shrimp Farming on Mangrove Ecosystems. CAB International, 3(003): 1-11.
- Avnimelech, Y. 1999. Carbon/Nitrogen Ratio as A Control Element in Aquaculture Systems. *Aquaculture*, 176: 227-235.
- Aziz, R., Nirmala, K., Affandi, R., dan Prihadi, T. 2015. Kelimpahan Plankton Penyebab Bau Lumpur Pada Budidaya Ikan Bandeng Menggunakan Pupuk N:P Berbeda. *Jurnal Akuakultur Indonesia*, 14(1): 58-68.
- Bassat, R. A. 2008. Composition and Abundance of The Zooplankton Community in The Bitter Lakes, Egypt, in Relation to Environmental Factors. *African Journal of Aquatic Science*, 33(3): 233-240.
- Bintari, N. W. D., Kawuri, R., and Dalem, A. A. G. R. 2016. Identifikasi Bakteri *Vibrio* Penyebab Vibriosis Pada Larva Udang Galah (*Macrobrachium rosenbergii* (De Man)). *Jurnal Biologi*, 20(2): 53-63.
- Boyd, C. E., and Tucker, C. S. 1998. Pond Aquaculture Water Quality Management. Kluwer academic Publishers. New York: 700 p.
- Cardenas, C. A. M., and Saavedra, M. P. S. 2017. Inhibitory Effect of Benthic Diatom Species on Three Aquaculture Pathogenic Vibrios. *Algal Research*, 27: 131-139.
- Chakravarty, M. S., Ganesh, P. R. C., Amarnath, D., Sudha, S., and Babu, T. S. 2016. Spatial Variation of Water Quality Parameters of Shrimp (*Litopenaeus vannamei*) Culture Ponds at Narsapurapupeta, Kajuluru and Kaikavolu Villages of East Godavari District, Andhra Pradesh. *International Journal of Fisheries and Aquatic Studies*, 4(4): 390-395.
- Chatterjee, S., and Haldar, S. 2012. *Vibrio* Related Diseases in Aquaculture and Development of Rapid and Accurate Identification Methods. *J. Marine Sci. Res. Dev.*, S1: 002.
- Culberson, S. D., and Piedrahita, R. H. 1996. Aquaculture Pond Ecosystem Model: Temperature and Dissolved Oxygen Prediction-Mechanism and Application. *Ecological Modelling*, 89:231-259.
- Daruti, D. N., Rozi., and Rahayu, K. 2017. The Effect of Hydrogen Peroxide on N/P Ratio and Phytoplankton Diversity in Vannamei Shrimp (*Litopenaeus vannamei*) Ponds in Banyuwangi, East Java. *IOP Conf. Series: Earth and Environmental Science*, 137: 1-7.
- Deep, P. R., Bhattacharyya, S., and Nayak, B. 2013. Cyanobacteria in Wetlands of The Industrialized Sambalpur District of India. *Aquatic Biosystems*, 9(14): 1-12.
- Directorate General of Aquaculture. 2016. Annual Performance Report. Ministry of Fisheries and Marine Affairs. 180 p.
- Edhy, W. A., Azhary, K., Pribadi, J., and Chaerudin, M. 2010. Budidaya Udang Putih (*Litopenaeus vannamei*. Boone, 1931). Jakarta: CV. Mulia Indah. 194 p.
- Fernandes, S. O., Kulkarni, S. S., Shiroadkar, R. R., Karekar, S. V., Kumar, R. P., Sreepada, R. A., Vogelsang, C., and Bharathi P. A. L. 2010. Water Quality and Bacteriology in An Aquaculture Facility Equipped with a New Aeration System. *Environmental Monitoring and Assessment*, 164(4): 81-92.
- Gopal, S., Otta, S. K., Kumar, S., Karunasagar, I., Nishibuchi, M., and Karunasagar, I. 2005. The Occurrence of *Vibrio* Species in

- Tropical Shrimp Culture Environments; Implications for Food Safety. *International Journal of Food Microbiology*, 102: 151-159.
- Gross, A., Abutbul, S., and Zilberg, D. 2004. Acute and Chronic Effects of Nitrite on White Shrimp, *Litopenaeus vannamei*, Cultured in Low-Salinity Brackish Water. *Journal of The World Aquaculture Society*, 35(3): 315-321.
- Han, Y. J., Jo, A., Kim, S. W., Lee, H. E., Kim, Y.C., Jeong, H. D., Choi, Y. H., Kim, S., Cha, H. J., and Kim, H. S., 2018. Multiplex PCR Using *YeaD* and 16S rRNA Gene to Identify Major Pathogens in Vibriosis of *Litopenaeus vannamei*, 41: 35-42.
- Hargreaves, J. A. 1998. Nitrogen Biogeochemistry of Aquaculture Ponds. *Aquaculture*, 166: 181-212.
- Harris, J. M., Vinobaba, P., Kularatne, R. K. A., and Champika, E. K. 2016. Spatial and Temporal Distribution of Cyanobacteria in Batticaloa Lagoon. *Journal of Environmental Sciences*, 47: 211-218 p.
- Hastuti, Y. P. 2011. Nitrifikasi dan Denitrifikasi di Tambak. *Jurnal Akuakultur Indonesia*, 10(1): 89-98.
- Hatmanti, A. 2003. Penyakit Bakterial Pada Budidaya Krustasea Serta Cara Penanganannya. *Oseana*, 28(3): 1-10.
- Heenatigala, P. P. M., and Fernando, M. U. L. 2016. Occurrence of Bacteria Species Responsible for Vibriosis in Shrimp Pond Culture Systems in Sri Lanka and Assessment of The Suitable Control Measures. *Sri lanka Journal Aquatic Science*, 21(1): 1-17.
- Hopkins, J. S., Sandifer, P. A., and Browdy, C. L. 1994. Sludge Management in Intensive Pond Culture of Shrimp: Effect of Management Regime on Water Quality, Sludge Characteristics, Nitrogen Extinction, and Shrimp Production. *Aquaculture Engineering*, 13: 11-30.
- Islam, M. S., Kamal, A. H. M. M., Wahab, M. A., and Dewan, S. 2004. Water Quality Parameters of Coastal Shrimp Farms from Southwest and Southeast Regions of Bangladesh. *Bangladesh Journal Fisheries Research*, 8(1): 53-60.
- Jaganmohan, P., and Kumari, C. L. 2018. Assessment of Water Quality in Shrimp (*L. Vannamei*) Grow Out Ponds in Selected Villages of S.P.S.R Nellore District of Andhra Pradesh, India During Winter Crop Season. *International Journal of Fisheries and Aquatic Studies*, 6(3): 260-266.
- Janeo, R. L., Corre, V. L., and Sakata, T. 2009. Water Quality and Phytoplankton Stability in Response to Application Frequency of Bioaugmentation Agent in Shrimp Ponds. *Aquaculture Engineering*, 40: 120-125.
- Jayanthi, M., Thirumurthy, S., Muralidhar, M., and Ravichandran, P. 2018. Impact of Shrimp Aquaculture Development on Important Ecosystems in India. *Global Environmental Change*, 52: 10-21.
- Jayasree, L., Janakiram, P., and Madhavi, R. 2006. Characterization of *Vibrio* spp. Associated with Diseased Shrimp from Culture Ponds of Andhra Pradesh (India). *Journal of The World Aquaculture Society*, 37(4): 523-532.
- Jescovitch, L. N., Ullman, C., Rhodes, M., and Davis, D. A. 2017. Effects of Different Feed Management Treatments on Water

- Quality for Pacific White Shrimp *Litopenaeus vannamei*. Aquaculture Research, 49(1): 1-6.
- Jia, W., Huang, X., and Li., C. 2014. A Preliminary Study of The Algicidal Mechanism of Bioactive Metabolites of *Brevibacillus Laterosporus* on *Oscillatoria* in Prawn Ponds. The Scientific World Journal, 2014: 1-11.
- Jiao, S., Maltecca, C., Gray, K. A., and Cassady, J. P. 2014. Feed Intake, Average Daily Gain, Feed Efficiency, and Real-Time Ultrasound Traits in Duroc Pigs: I. Genetic Parameter Estimation and Accuracy of Genomic Prediction. Journal of Animal Science, 92: 2377-2386.
- Junda, M. 2018. Development of Intensive Shrimp Farming, *Litopenaeus vannamei* In Land-Based Ponds: Production and Management. Journal of Physics: Conf. Series, 1029: 1-6.
- Kharisma, A., dan Manan, A. 2012. Kelimpahan Bakteri *Vibrio* sp. Pada Air Pembesaran Udang Vannamei (*Litopenaeus vannamei*) Sebagai Deteksi Dini Serangan Penyakit Vibriosis. Jurnal Ilmiah Perikanan dan Kelautan, 4(2): 129-134.
- Kopprio, G. A., Streitenberger, M. E., Okuno, K., Baldini, M., Biancalana, F., Fricke, A., Martinez, A., Neogi, S. B., Koch, B. P., Yamasaki, S., and Lara, R. J. 2017. Biogeochemical and Hydrological Drivers of The Dynamics of *Vibrio* Species in Two Patagonian Estuaries. Science of The Total Environment, 579: 646-656.
- Lekshmi, S., Nansimole, A., Mini, M., Athira, N., and Radhakrishnan, T., 2014. Occurrence of *Vibrio Cholerae* in Shrimp Culture Environments of Kerala, India. Indian Journal Science and Research, 5(2): 151-160.
- Lovell, R. T., and Broce, D. 1985. Cause of Musty Flavor in Pond-Cultured Penaeid Shrimp. Aquaculture, 50: 169-174.
- Mangampa, M., 2015. Dinamika Populasi Bakteri dalam Air dan Sedimen Tambak pada Pemantapan Budidaya Udang Vaname Ekstensif Plus Melalui Pergiliran Pakan. Berkala Perikanan Terubuk, 43(2): 25-35.
- Masithah, E. D., Octaviana, Y. D., and Manan, A. 2016. Pengaruh Perbedaan Probiotik Komersial Terhadap Rasio C:N dan N:P Media Kultur Bioflok Pada Bak Percobaan. Journal of Aquaculture and Fish Health, 5(3): 118-125.
- Mastan, C. A., 2015. Incidences of White Feces Syndrome (WFS) in Farm-Reared Shrimp, *Litopenaeus vannamei*, Andhra Pradesh. Indo American Journal of Pharmaceutical Research, 5(9): 3044-3047.
- Mateo, P., Leganes, F., Perona, E., Loza, V., and Pinas, F. F. 2015. Cyanobacteria as Bioindicators and Bioreporters of Environmental Analysis in Aquatic Ecosystems. Biodiversity and Conservation, 24: 909-948.
- Michael, P. 1995. Metode Ekologi Untuk Penyelidikan Ladang dan Laboratorium. Universitas Indonesia. Jakarta.
- Minister of Marine and Fisheries of the Republic Indonesia No. 75 2016 th, 2016. General Guidelines for Cultivation of Black Tiger Shrimp (*Penaeus monodon*) and White Leg Shrimp (*Litopenaeus vannamei*). Jakarta: 43 p.

- Mohan, K., Ravichandran, S., Muralisankar, T., Uthayakumar, V., Chandirasekar, R., Seedeve, P., Rajan, D. K. 2019. Potential Uses of Fungal Polysaccharides as Immunostimulants in Fish and Shrimp Aquaculture: A Review. *Aquaculture*, 500: 250-263.
- Nimrat, S., Suksawat, S., Maleeweatch, P., and Vuthiphandchai, V. 2008. Effect of Different Shrimp Pond Bottom Soil Treatments on The Change of Physical Characteristics and Pathogenic Bacteria in Pond Bottom Soil. *Aquaculture*, 285: 123-129.
- Nurhatijah, N., Muchlisin, Z. A., Sarong, M. A., and Supriatna, A. 2016. Application of Biofloc to Maintain the Water Quality in Culture System of The Tiger Prawn (*Penaeus monodon*). *AACL Bioflux*, 9(4): 923-928.
- Orozco, L. N., Felix, E. A., Ciapara, I. H., Flores, R. J., and Cano, R. 2007. Pathogenic and Non-Pathogenic *Vibrio* Species in Aquaculture Shrimp Ponds. *Revista Latinoamericana de Microbiologia*, 49(3-4): 60-67.
- Palafox, J. T. P., Figueroa, J. L. A., Vargasmachuca, S. G. C., Chavez, G. R., Valle, B. A., Dios, M. A. R., Carrillo, F. M., Villalobos, R. N., Gurrola, J. A. G., and Lugo, P. L. 2010. The Effect of Chemical and Organic Fertilization on Phytoplankton and Fish Production in Carp (*Ciprinidae*) Polyculture System. *Revista Biociencias*, 1(1): 44-50.
- Palafox, J. T. P., Pavia, A. A., Lopez, D. G. M., Figueroa, J. L. A., Reynoso, F. L., Chavez, M. R. C., Leal, H. E., Luna, A. R., Ozuna, F. P., Vargasmachuca, S. G. C., Gomez, V. P. 2019. Response Surface Analysis of Temperature-Salinity Interaction Effects on Water Quality, Growth and Survival of Shrimp *Penaeus vannamei* Postlarvae Raised in Biofloc Intensive Nursery Production. *Aquaculture*, 503: 312-321.
- Panjaitan, P., 2011. Effect of C: N Ratio Levels on Water Quality and Shrimp Production Parameters in *Penaeus monodon* Shrimp Culture with Limited Water Exchange Using Molasses as a Carbon Source. *Ilmu Kelautan*. Vol. 16(1): 1-8 p.
- Pirzan, A. M., dan Masak, P. R. P. 2007. Hubungan Produktifitas Tambak Dengan Keragaman Fitoplankton di Sulawesi Selatan. *Jurnal Riset Akuakultur*, 2(2): 211-220.
- Prescott, L. M., Harley, J. P., Klein, O. A. 2002. Human Diseases Caused by Bacteria. In: *Microbiology*, 5<sup>th</sup> ed. Prescott, L. M., Harley, J. P., Klein, O. A. (eds). Mc Graw-Hill Publishers: 732-735 p.
- Rahman, M. Z., Zaman, M. FU., Khodoker, S., Jaman, M. H. U., Hossain, M. L., Bappa, S. B. 2015. Water Quality Assessment of a Shrimp Farm: A Study in A Salinity Prone Area of Bangladesh. *International Journal of Fisheries and Aquatic Studies*, 2(5): 9-19.
- Redfield, A. C., Ketchum, B. H., and Richards, F. A. 1963. The Influence of Organism on The Composition of Water. In: Hill, M. N. (Ed.) *The Sea* v.2, New York: 26-77 p.
- Rimi, R. H., Farzana, S., Sheikh, M. S., Abedin, M. Z, and Bhowmick, A. C. 2013. Climate Change Impacts on Shrimp Production at the South-West Coastal Region of Bangladesh. *World Environment*, 3(3): 116-125.
- Rodgers, J. H. 2008. *Algal Toxins in Pond Aquaculture*. SRAC Publication. No. 4605.

- Rochin, J. R., Espericueta, M. G. F., Sanudo, J. F. F., Silvas, S. G. A., Lopez, M. G. F., and Osuna, F. P. 2017. Acute Toxicity of Nitrite on White Shrimp *Litopenaeus vannamei* (Boone) Juveniles in Low-Salinity Water. *Aquaculture Research*, 48: 2337-2343.
- Romadhona, B., Yulianto, B., Sudarno. 2016. Fluktuasi Kandungan Amonia dan Beban Cemar Lingkungan Tambak Udang Vaname Intensif Dengan Teknik Panen Parsial dan Panen Total. *Jurnal Saintek Perikanan*, 11(2): 84-93.
- Sarjito, Apriliani, M., Afriani, D., dan Haditomo, A. H. C. 2015. Agensia Penyebab Vibriosis Pada Udang Vaname (*Litopenaus vannamei*) Yang Dibudidayakan Secara Intensif Di Kendal. *Jurnal Kelautan Tropis*, 18(3): 189-196.
- Schober, J., Lima, G., and Focken, U. 2007. Analysis of Soil Nutrients and Organic Matter in Organic and Conventional Marine Shrimp Ponds at Guaraíra Lagoon, Rio Grande do Norte State, Brazil. *Wissenschaftstagung Ökologischer Landbau*: 1-5.
- Singh, R. R., Raja, K., Gopalakrishnan, A., Kannan, D., and Sakthivel, A. 2013. Study on Growth Performance of *Litopenaeus vannamei* Cultured in Bore Well and Brackish Water Fed Ponds. *International Journal of Current Research*, 5(11): 3567-3570.
- Smith, P. T. 1996. Toxic Effects of Blooms of Marine Species of Oscillatoriales on Farmed Prawns (*Penaeus monodon*, *Penaeus japonicus*) and Brine Shrimp (*Artemia salina*). *Toxicom*, 34(8): 857-869.
- Soto, D., Ross, L. G., Handisyde, N., Bueno, P. B., Beveridge, M. C. M., Dabbadie, L., Manjarrez, J. A., Cai, J., and Pongthanapanich, T. 2018. Climate Change and Aquaculture: Vulnerability and Adaptation Options. Food and Agriculture Organization of The United Nations. Roma: 465-490 p.
- Sriurairatana, S., V. Boonyawiwat., W. Gangnonngiw., C. Laosutthipong., J. Hiranchan and T. W. Flegel. 2014. White Feces Syndrome of Shrimp Arises from Transformation, Sloughing and Aggregation of Hepatopancreatic Microvilli into Vermiform Bodies Superficially Resembling Gregarines. *Plos One*, 9 (6): 1-8.
- Stalin, N., and Srinivasan, P. 2017. Efficacy of Potential Phage Cocktails Against *Vibrio* *Harveyi* and Closely Related *Vibrio* Species Isolated from Shrimp Aquaculture Environment in The South East Coast of India. *Veterinary Microbiology*, 207: 83-96.
- Supono. 2015. Manajemen Lingkungan Untuk Akuakultur. Plantaxia. Yogyakarta: 116 hal.
- Suwanpakdee, S., Direkbusarakom, S., Chotipuntu, P., and Songsangjinda, P. 2010. Urea as a Nitrogen Source in a Black Tiger Shrimp (*Penaeus monodon*) Closed Culture System. *Walailak Journal Science and Technology*, 7(2): 135-140.
- Syah, R., Makmur., Fahrur, M., 2017. Budidaya Udang Vaname Dengan Padat Penebaran Tinggi. *Media Akuakultur*. Vol. 12(1): 19-26 hal.
- Tarunamulia, Kamariah, dan Mustafa, A. 2016. Keterkaitan Spasial Kualitas Lingkungan dan Keberadaan Fitoplankton Berpotensi HABs Pada Tambak Ekstensif di

- Kecamatan Losari Kabupaten Cirebon, Jawa Barat. Jurnal Riset Akuakultur, 11(2): 181-195.
- Thakur, D. P., and Lin., C. K. 2003. Water Quality and Nutrient Budget in Closed Shrimp (*Penaeus monodon*) Culture Systems. Aquaculture Engineering, 27: 159-176.
- Tho, N., Merckx, R., and Ut, V.N. 2012. Biological Characteristics of The Improved Extensive Shrimp System in The Mekong Delta of Vietnam. Aquaculture Research, 43: 526-537.
- Tompo, A. 2016. Kajian Populasi Bakteri *Vibrio* sp. Pada Tambak Budidaya Udang Vaname (*Litopenaeus vannamei*) Sistim Semi Intensif Dengan Persentase Pemberian Pakan Yang Berbeda. Jurnal Ilmu Perikanan, 5(1): 470-475.
- Tucker, C. S. 2000. Off\_Flavor Problems in Aquaculture. Reviews in Fisheries Science, 8(1): 45-88.
- Utami, W., Sarjito, dan Desrina. 2016. Pengaruh Salinitas Terhadap Efek Infeksi *Vibrio harveyi* Pada Udang Vaname (*Litopenaeus vannamei*). Journal of Aquaculture Management and Technology, 5(1): 82-90.
- Utojo. 2015. Keragaman Plankton dan Kondisi Perairan Tambak Intensif dan Tradisional di Probolinggo Jawa Timur. Biosfera, 32(2): 83-97.
- Weidemann, G. 2002. Control of Blue-Green Algae in Aquaculture Ponds. SRAC International: 20-49 p.
- Widigdo, B., dan Wardiatno, Y. 2013. Dinamika Komunitas Fitoplankton dan Kualitas Perairan Di Lingkungan Perairan Tambak Udang Intensif: Sebuah Analisis Korelasi. Jurnal Biologi Tropis, 13(2): 160-184.
- Xu, H., Paerl, H. W., Qin, B., Zhu, G., and Gao, G. 2010. Nitrogen and Phosphorus Inputs Control Phytoplankton Growth in Eutrophic Lake Taihu, China. Limnology Oceanography, 55(1): 420-432.
- Yi, D., Reardon, T., Stringer, R., 2016. Shrimp Aquaculture Technology Change in Indonesia: Are Small Farmers Included? Aquaculture, 493: 436-445.
- Yusoff, F. M., Matias, H. B., Khalid, Z. A., and Phang, S. M. 2011. Culture of Microalgae Using Interstitial Water Extracted from Shrimp Pond Bottom Sediments. Aquaculture. Vol. 201: 263-270 p.
- Zafar, M. A., Haque, M. M., Aziz, M. S. B., and Alam, M. M. 2015. Study on Water and Soil Quality Parameters of Shrimp and Prawn Farming in The Southwest Region of Bangladesh. J. Bangladesh Agril. Univ., 13(1): 153–160.